Insecticide sprays, natural enemy assemblages and predation on Asian citrus psyllid, *Diaphorina citri* (Hemiptera: Psyllidae)

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Abstract

The Asian citrus psyllid (ACP), *Diaphorina citri* Kuwayama is considered a key citrus pest due to its role as vector of ‘huanglongbing’ (HLB) or citrus greening, probably the most economically damaging disease of citrus. Insecticidal control of the vector is still considered a cornerstone of HLB management to prevent infection and to reduce reinoculation of infected trees. The severity of HLB has driven implementation of intensive insecticide programs against ACP with unknown side effects on beneficial arthropod fauna in citrus agroecosystems. We evaluated effects of calendar sprays directed against this pest on natural enemy assemblages and used exclusion to estimate mortality they imposed on ACP populations in citrus groves. Predator exclusion techniques were used on nascent colonies of *D. citri* in replicated large untreated and sprayed plots of citrus during the four major flushing periods over 2 years. Population of spiders, arboreal ants and ladybeetles were independently assessed. Monthly sprays of recommended insecticides for control of ACP, adversely affected natural enemy populations resulting in reduced predation on ACP immature stages, especially during the critical late winter/early spring flush. Consequently, projected growth rates of the ACP population were greatest where natural enemies had been adversely affected by insecticides. Whereas, this result does not obviate the need for insecticidal control of ACP, it does indicate that even a selective regimen of sprays can impose as yet undetermined costs in terms of reduced biological control of this and probably other citrus pests.

Keywords: conservation biological control, huanglongbing, population growth rate, predator exclusion, side effects

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Introduction

The Asian citrus psyllid (ACP), *Diaphorina citri* Kuwayama is a key pest of citrus in Asia and the Americas, due primarily to its role as vector of ‘huanglongbing’ (HLB) or citrus greening disease (Grafton-Cardwell et al., 2013). The causal agent of HLB in these regions is considered to be Candidatus Liberibacter asiaticus, a phloem-limited Gram-negative bacterium. HLB reduces tree health, productivity and fruit quality. Infected trees without intervention decline and become unproductive within 5–10 years (Bove, 2006).

First detections of HLB in America occurred in Brazil in 2004 and a year later in Florida (Halbert, 2005; Belasque et al., 2010) raising the status of ACP to key pest in the two main citrus producing areas of the world. By 2012, the disease had spread throughout most of the Caribbean region and Mexico with detections in Texas and California. A recent study...
estimated total economic impact of $4.54 billion in Florida from 2006 to 2011, including loss of 8257 jobs (Hodges & Sreen, 2012).

Host plant resistance is expected to provide the ultimate long-term solution to HLB. However, with nothing yet on the immediate horizon, short-term measures are needed to maintain economic production levels in endemic areas. Vector control is considered essential, along with roguing of symptomatic trees (Bove, 2006) or therapeutic applications of foliar nutrients to mitigate effects of the disease in high incidence areas (Stansly et al., 2013). The challenge with vector control is integrating insecticide programs with the potentially important component of vector suppression offered by beneficial insects and mites (Michaud, 2004; Qureshi & Stansly, 2009).

Insecticidal control of the vector still is an important component of HLB management (Qureshi & Stansly, 2008, 2010; Rogers et al., 2012). However, fear and severity of HLB have driven implementation of intensive insecticide programs to control the psyllid vector (Belasque et al., 2010; Rogers et al., 2012), that could exert important side effects against beneficial arthropod fauna in citrus agroecosystems (Qureshi & Stansly, 2007). Consequently, insecticidal control strategies that maximize ACP control while maintaining natural enemy diversity and ecological stability are being developed and promoted (Qureshi & Stansly, 2010). These strategies include foliar sprays of broad-spectrum insecticides applied during the dormant period of plant growth, or at the end of the growing season when natural enemy activity declines, sampling techniques to monitor ACP populations, and guides for the use of selective insecticides during the growing season (Hall et al., 2007; Qureshi & Stansly, 2008, 2010).

_D. citri_ requires tender plant tissue to complete its life cycle in citrus. Adult females lay eggs on new growth shoots (referred to as flush) upon which all nymphal stages develop (Shivanker et al., 2000). Plant phenology is therefore a key factor influencing _D. citri_ life history and seasonal demography. Major flushing periods during the growing season provide ACP with abundant resources for reproduction. Biological control mainly affects pre-imago stages which are exclusively on new flush. Therefore, the regulatory role exerted by biological control on pest demography in citrus would normally be greatest during periods of new foliage growth.

The principal parasitoid attacking _D. citri_ is _Tamarixia radiata_ Waterston (Hymenoptera: Eulophidae), established throughout Florida but with limited impact on ACP, due presumably to insecticide use and low survival in winter (Qureshi et al., 2009). Programs are in place to evaluate the effectiveness of augmentation, especially in spring when parasitism levels tend to be lowest (Qureshi et al., 2009). In contrast, certain naturally occurring species of ladybeetle (Coleoptera: Coccinellidae), lacewings (Neuroptera: Chrysopidae), spiders (Araneae) and other generalist predators have been seen to inflict up to 100% mortality on cohorts of _D. citri_ nymphs during the growing season in Florida (Michaud, 2004; Qureshi & Stansly, 2009). These and several other natural enemies, including predaceous arboreal ants of the genus _Pseudomyrmex_, also suppress and prevent a wide range of citrus pests in Florida from causing economic damage (McCoy et al., 2009). Given that naturally occurring biological control provides a valuable service, it is paramount to document costs incurred when beneficial arthropods are suppressed by pesticides, specifically impacts derived from intensified insecticidal spray programs targeting ACP.

We evaluated populations of key predator guilds in large untreated citrus blocks compared to blocks subjected to monthly sprays of selective and broad-spectrum insecticides targeting ACP. We also used predator exclusion techniques on nascent colonies of immature ACP in these blocks to estimate the impact of foliar sprays on ACP biological control and natural enemies. The insecticide strategy followed in the treated plots was designed to minimize pesticide resistance and side effects on beneficial arthropod fauna. Our hypothesis was that continuous insecticide applications for ACP control, even using selective pesticides, may have deleterious and cumulative effects on biological control of this pest. The information obtained could be used to discern ways to improve management programs that optimize ACP control, in part by reaping maximum benefit from pest suppression provided by predaceous insects.

### Material and methods

#### Study site and experimental design

Experiments were conducted in a 10.3ha commercial citrus grove located near LaBelle (Hendry County, FL, USA) (26°41’04"N, 81°26’20"W). The grove was planted December 2001 with sweet orange, _Citrus sinensis_ (L.) Obseck ‘Early Gold’, bud-grafted to ‘Carrizo’ cirtage rootstock at a density of 231 trees ha^-1_. Trees were irrigated by micro-sprinklers and conventional cultural practices were followed (Jackson, 1999). However, foliar nutrition was intensified with a HLB nutritional remediation program consisting sprays of 1.24kg ha^-1_ N; 7.73kg ha^-1_ K₂O; 2.14kg ha^-1_ P₂O₅; 1.11kg ha^-1_ [Zn]²⁺; 0.94kg ha^-1_ [Mg]²⁺; 3.05kg ha^-1_ [Mn]²⁺; 59.54kg ha^-1_ [Na]₂MoO₄; and 0.77kg ha^-1_ [B]³⁺ during the major flushing periods of the year (Stansly et al., 2013). Two ACP management treatments: calendar sprays of insecticides to control ACP (‘calendar’) and untreated control (‘no insecticide’) were tested in a randomized complete block design with four replications. The ‘calendar’ treatment was aimed at maintaining ACP densities as low as possible while still conserving natural enemies by generally avoiding broad-spectrum insecticides during the growing season. Plot dimensions (length × width) were 38 × 170 m containing approximately 144 mature trees and several resets. Treated and untreated plots were randomly distributed through the 10.3ha commercial citrus grove. Minimum distance between sampling areas in untreated plots and treated plots was of 27 m in the case of adjacent plots and 62 m in the case that a treated and an untreated plot were not adjacent.

#### Pest and disease management

Monthly insecticide applications directed against ACP were initiated in July 2010 (table 1). Broad-spectrum products (organophosphates, carbamates and pyrethroids) were generally restricted to the winter and the end of the summer, and more selective insecticides were used preferentially during the growing season. Nine different groups of insecticides were selected and rotated to avoid inducing resistance in _D. citri_ to any particular one. The untreated control was not sprayed for ACP. The entire block was sprayed three times in 2011 and two times in 2012 with copper-based products to control citrus canker, _Xanthomonas axonopodis pv._ citri. In addition, a liquid
Table 1. Spray application dates, products, % active ingredient, rates, objective, treatments included and schedule of exclusion experiments conducted in a commercial sweet orange citrus grove. Treatments: calendar Asian citrus psyllid (ACP) insecticide applications (1), untreated (2). ‘Nutrients’ refers to foliar nutritional remediation sprays applied to mitigate deficiencies associated to huanglongbing (HLB).

<table>
<thead>
<tr>
<th>Dates</th>
<th>Active ingredient (brand name and formulation)</th>
<th>% Active ingredient</th>
<th>Rate</th>
<th>Objective</th>
<th>Treatments sprayed</th>
<th>Exclusion experiments</th>
</tr>
</thead>
<tbody>
<tr>
<td>30 July 2010</td>
<td>Spinetoram (Delegate WG)</td>
<td>25</td>
<td>315 g ha$^{-1}$</td>
<td>ACP control</td>
<td>1</td>
<td>x</td>
</tr>
<tr>
<td>13 October 2010</td>
<td>Dimethoate (Dimethoate 4E)</td>
<td>43.5</td>
<td>1.171 g ha$^{-1}$</td>
<td>ACP control</td>
<td>1</td>
<td>x</td>
</tr>
<tr>
<td>20 January 2011</td>
<td>Fenpropathrin (Danitol 2.4 EC)</td>
<td>30.9</td>
<td>0.581 g ha$^{-1}$</td>
<td>ACP control</td>
<td>1</td>
<td>x</td>
</tr>
<tr>
<td>4 March 2011</td>
<td>Nutrients + copper (Kocide 3000) ***</td>
<td>***</td>
<td>***</td>
<td>HLB remediation + Canker control</td>
<td>1, 2</td>
<td>x</td>
</tr>
<tr>
<td>16 March 2011</td>
<td>Diflubenzuron (Micromite 80WGS)</td>
<td>80</td>
<td>438 g ha$^{-1}$</td>
<td>ACP control</td>
<td>1</td>
<td>x</td>
</tr>
<tr>
<td>15 April 2011</td>
<td>Carbaryl (Sevin XLR Plus)</td>
<td>44.1</td>
<td>7.021 g ha$^{-1}$</td>
<td>ACP control</td>
<td>1</td>
<td>x</td>
</tr>
<tr>
<td>18 May 2011</td>
<td>Spinetoram (Delegate WG)</td>
<td>25</td>
<td>315 g ha$^{-1}$</td>
<td>ACP control</td>
<td>1</td>
<td>x</td>
</tr>
<tr>
<td>15 June 2011</td>
<td>Imidacloprid (Admire Pro)</td>
<td>42.8</td>
<td>315 g ha$^{-1}$</td>
<td>ACP control</td>
<td>1</td>
<td>x</td>
</tr>
<tr>
<td>15 July 2011</td>
<td>Nutrients ***</td>
<td>***</td>
<td>***</td>
<td>HLB remediation</td>
<td>1, 2</td>
<td>x</td>
</tr>
<tr>
<td>28 July 2011</td>
<td>Abamectine (Agri-Mek SC)</td>
<td>8</td>
<td>0.261 g ha$^{-1}$</td>
<td>ACP control</td>
<td>1</td>
<td>x</td>
</tr>
<tr>
<td>1 August 2011</td>
<td>Sulfur 80% (Microthiol Disperss)</td>
<td>80</td>
<td>17 kg ha$^{-1}$</td>
<td>Rust mite control</td>
<td>1, 2</td>
<td>x</td>
</tr>
<tr>
<td>9 August 2011</td>
<td>Nutrients + copper (Cuprofix Ultra 40D) ***</td>
<td>***</td>
<td>***</td>
<td>HLB remediation + Canker control</td>
<td>1, 2</td>
<td>x</td>
</tr>
<tr>
<td>19 August 2011</td>
<td>Malathion (Gowan Malathion 8F)</td>
<td>79.5</td>
<td>2.921 g ha$^{-1}$</td>
<td>ACP control</td>
<td>1</td>
<td>x</td>
</tr>
<tr>
<td>16 September 2011</td>
<td>Fenpropatrin (Danitol 2.4 EC)</td>
<td>30.9</td>
<td>0.581 g ha$^{-1}$</td>
<td>ACP control</td>
<td>1</td>
<td>x</td>
</tr>
<tr>
<td>30 September 2011</td>
<td>Nutrients + copper (Cuprofix Ultra 40D) ***</td>
<td>***</td>
<td>***</td>
<td>HLB remediation + Canker control</td>
<td>1, 2</td>
<td>x</td>
</tr>
<tr>
<td>2 November 2011</td>
<td>Spirotetramat (Movento MPC)</td>
<td>14.5</td>
<td>1.171 g ha$^{-1}$</td>
<td>ACP control</td>
<td>1</td>
<td>x</td>
</tr>
<tr>
<td>2 December 2011</td>
<td>Carbaryl (Sevin XLR Plus)</td>
<td>44.1</td>
<td>7.021 g ha$^{-1}$</td>
<td>ACP control</td>
<td>1</td>
<td>x</td>
</tr>
<tr>
<td>19 December 2011</td>
<td>Phosmet (Imidan 70-W)</td>
<td>70</td>
<td>1122 g ha$^{-1}$</td>
<td>ACP control</td>
<td>1</td>
<td>x</td>
</tr>
<tr>
<td>12 January 2012</td>
<td>Zeta-cypermethrin (Mustang 1.5 EW)</td>
<td>17.1</td>
<td>301 g ha$^{-1}$</td>
<td>ACP control</td>
<td>1</td>
<td>x</td>
</tr>
<tr>
<td>29 February 2012</td>
<td>Nutrients ***</td>
<td>***</td>
<td>***</td>
<td>HLB remediation</td>
<td>1, 2</td>
<td>x</td>
</tr>
<tr>
<td>7 March 2012</td>
<td>Spirotetramat (Movento MPC)</td>
<td>14.5</td>
<td>1.171 g ha$^{-1}$</td>
<td>ACP control</td>
<td>1</td>
<td>x</td>
</tr>
<tr>
<td>26 April 2012</td>
<td>Diflubenzuron (Micromite 80WGS)</td>
<td>80</td>
<td>438 g ha$^{-1}$</td>
<td>ACP control</td>
<td>1</td>
<td>x</td>
</tr>
<tr>
<td>1 May 2012</td>
<td>Nutrients + copper (Kocide 3000) ***</td>
<td>***</td>
<td>***</td>
<td>HLB remediation + Canker control</td>
<td>1, 2</td>
<td>x</td>
</tr>
<tr>
<td>6 June 2012</td>
<td>Abamectine (Abbacus) + nutrients</td>
<td>8</td>
<td>0.261 g ha$^{-1}$</td>
<td>HLB remediation + rust mite control</td>
<td>1, 2</td>
<td>x</td>
</tr>
<tr>
<td>29 June 2012</td>
<td>Spirotetramat (Delegate WG)</td>
<td>25</td>
<td>315 g ha$^{-1}$</td>
<td>ACP control</td>
<td>1</td>
<td>x</td>
</tr>
<tr>
<td>23 July 2012</td>
<td>Nutrients + copper (Cuprofix Ultra 40D) ***</td>
<td>***</td>
<td>***</td>
<td>HLB remediation + Canker control</td>
<td>1, 2</td>
<td>x</td>
</tr>
<tr>
<td>10 August 2012</td>
<td>Imidacloprid (Admire Pro)</td>
<td>42.8</td>
<td>315 g ha$^{-1}$</td>
<td>ACP control</td>
<td>1</td>
<td>x</td>
</tr>
<tr>
<td>13 August 2012</td>
<td>Nutrients ***</td>
<td>***</td>
<td>***</td>
<td>HLB remediation</td>
<td>1, 2</td>
<td>x</td>
</tr>
<tr>
<td>18 September 2012</td>
<td>Dimethoate (Dimethoate 4E)</td>
<td>43.5</td>
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<td>ACP control</td>
<td>1</td>
<td>x</td>
</tr>
<tr>
<td>1 October 2012</td>
<td>Sulfur 80% (Microthiol Disperss)</td>
<td>80</td>
<td>17 kg ha$^{-1}$</td>
<td>Rust mite control</td>
<td>1, 2</td>
<td>x</td>
</tr>
<tr>
<td>19 October 2012</td>
<td>Fenpyroximate (Portal)</td>
<td>5</td>
<td>4.681 g ha$^{-1}$</td>
<td>ACP control</td>
<td>1</td>
<td>x</td>
</tr>
<tr>
<td>17 December 2012</td>
<td>Zeta-cypermethrin (Mustang 1.5 EW)</td>
<td>17.1</td>
<td>301 g ha$^{-1}$</td>
<td>ACP control</td>
<td>1</td>
<td>x</td>
</tr>
<tr>
<td>11 January 2013</td>
<td>Phosmet (Imidan 70-W)</td>
<td>70</td>
<td>1122 g ha$^{-1}$</td>
<td>ACP control</td>
<td>1</td>
<td>x</td>
</tr>
</tbody>
</table>

*** indicate that no active ingredient % and Rate are provided in the table for nutrients.

‘x’ under ‘Exclusion experiments’ indicates when the exclusion experiments were conducted with respect to the sprays dates provided in the first column ‘Dates’.
sulfur formulation was applied once in 2011 and 2012 and abamectin in 2012 over the whole block to control citrus rust mite, *Phyllocoptruta oleivora* (Ashmead) (Acari: Eriophyidae).

**Citrus flush density**

Density of emerging shoots (flush) suitable for colonization by ACP was estimated every 2 weeks from 4 April 2011 to 3 April 2013 in all plots by counting the total number of trees needed to find ten new shoots in an approximately 1.5 m × 1.5 m area of the canopy of 20 trees. When more than three new shoots were found in the 150 cm × 150 cm area of the first four trees, a 0.8 m² PVC (Polyvinyl chloride) square was randomly placed over the canopy of 12 trees per plot, and the total number of new shoots falling within the square was counted. Daily mean temperatures and rainfall for the same period of time from the University of Florida weather station in Immokalee, 25 km to the south were used to interpret tree phenology data.

**Monitoring ACP adults**

Population of ACP adults were monitored approximately 2, 4 and 6 weeks before and after each major flushing period by conducting two stem tap samples on two sides of 20 randomly selected trees in each plot. Adults were counted that fell on a clipboard covered with a 22 × 28 cm laminated white sheet held horizontally under a randomly chosen branch struck three times with a length of PVC pipe to make one ‘tap’ sample (Qureshi & Stansly, 2007).

**Monitoring natural enemies**

Population of natural enemies were assessed every 2 weeks from March 2011 to March 2013 using the stem tap samples described above. Spiders (Araneae), arboreal ants (Hymenoptera: Formicidae, Pseudomyrmecinae), lady beetles (Coleoptera: Coccinellidae) and lacewings (Neuroptera) previously identified as key natural enemies of ACP or other important citrus pests by Michaud (2001, 2004), Qureshi & Stansly (2009) and Xiao et al. (2007) were counted. Data were pooled according to meteorological seasons (spring: March, April, May; summer: June, July and August; autumn: September, October and November; winter: December, January and February) and cumulative numbers for each predatory group and season were calculated.

**Exclusion experiments**

Exclusion experiments were repeated eight times over a 2-year period, and initiated 25 May, 6 July, 23 August 2011 and 10 February, 10 May, 10 July, 9 September 2012 and 1 February 2013 or twice during each major flushing period. Synchronous cohorts of *D. citri* nymphs were established for each experiment by first caging each emerging shoot suitable for ACP oviposition with six ACP adults obtained from a greenhouse colony maintained on orange jasmine *Murraya paniculata* since 2005. A total of 72 cages, nine per plot, were set out each experiment. Exclusion cages were made of transparent polyethylene terephthalate cylindrical plastic jars 10.5 cm in diameter and 25 cm in height (CPS Inc. ID, USA). Top and bottom sections of the jars were removed and six additional 9.2 cm × 9.2 cm windows were cut in the sides. The resulting frames were all initially covered in a fine mesh organandy sleeve and fixed to the stem branch by three wires (30 cm long and 1.5 mm diameter) attached to the base of the plastic frame (fig. 1a). *D. citri* adults were removed from the cages after a 3-day oviposition period (4-day in February experiments) and branches containing shoots that had between 20 and 60 ACP eggs (34.6 ± 0.8) were selected for the experiments.

Four open and four closed cages were randomly assigned to the established colonies in ‘calendar’ or ‘no insecticide’ plots for a maximum of 64 colonies across four replicates (eight per exclusion treatment and plot). Closed cages were as above but open cages had circular holes cut in the organandy sleeve corresponding to the six windows in the frame and representing 26.4% of the total area of the cage to allow access by predator insects and mites (fig. 1b). All cages were removed from colonies after 7 days by which time most of the predation has occurred (Michaud, 2004; Qureshi & Stansly, 2009). Caged shoots hosting colonies were taken to the laboratory and the number of live *D. citri* nymphs remaining on each counted under a stereoscopic microscope.

**Statistical analysis**

Similarities in seasonal flush density patterns between ‘calendar’ and ‘no insecticide’ plots were analyzed using Spearman rank correlation. Differences in beneficial arthropod
abundance between ACP management treatments for each meteorological season and differences in the seasonal abundance pattern for each natural enemy group and between ‘calendar’ and ‘no insecticide’ treatments were studied by linear mixed model repeated measures analysis, where ACP management treatment was considered as a fixed factor and ‘block’ as random factor. Several covariance structures were tested and a heterogeneous autoregressive structure was selected for spiders, arboreal ants and lacewings based on the Akaike and Bayesian information criteria, respectively. Both criteria favor models that maximize goodness of fit while minimizing number of parameters. Ladybeetle data did not meet normality and homoscedasticity assumptions so the Kruskal–Wallis non-parametric test was used to determine seasonal and treatment effects on cumulative numbers.

Nymphs recovered at the end of each experiment were compared against the initial number of eggs in each colony in open or closed cages to calculate reduction in numbers. Percentage mortality in open cages corrected for mortality in closed cages was estimated for each plot using the Henderson and Tilton formula (Henderson, 1955).

\[
\left(1 - \frac{\text{eggs in exclusion} \times \text{nymphs in no exclusion}}{\text{nymphs in exclusion} \times \text{eggs in no exclusion}}\right) \times 100
\]

Corrected mortality attributed to predation was then compared between ‘calendar’ and ‘no insecticide’ treatments using linear mixed model repeated measures analysis. Changes in nymphal biotic reduction throughout the season as well as differences in seasonal biotic reduction patterns between the two ACP management treatments were also evaluated. A heterogeneous Toeplitz covariance structure was selected for analysis by the criteria mentioned above.

The effect of presumed predation during each major flushing period on ACP population growth rate (\(\lambda\)) was estimated by comparing stem tap results from ‘no insecticide’ plots before and after each flushing period \((n=3)\) using

\[
\Delta\text{ACP}^\# = \lambda = \frac{\text{ACP}^\# \text{ after} - \text{ACP}^\# \text{ before}}{\text{ACP}^\# \text{ before}}
\]

Values of \(\lambda\) would be positive if numbers increased over the interval, negative if they decreased and 0 if remained the same. Results for each of the eight major flushing periods were correlated using non-linear least-squares regression to the corresponding corrected mortality values \((n)\) for each of the eight major flushing periods evaluated. Data were fitted to a negative exponential equation \((3)\) by using the Newton–Raphson iterative estimation procedure

\[
\lambda = ab^{-r} + c
\]

Results

Citrus flush density

Four major flushing periods (more than 5 flushes m\(^{-2}\)) were registered between April 2011 and April 2013 with similar patterns for both treatments (Spearman’s \(\rho_{48} = -0.96, P<0.0001\)). The greatest flush density was observed in the first major flushing period that followed the coolest and driest period of the year (fig. 2). In 2012, the first major flushing period occurred in February (338.5 ± 10.3 flushes m\(^{-2}\)) whereas in 2013 it began earlier, in January (264.1 ± 7.2 flushes m\(^{-2}\)) and was more prolonged, following unseasonable mild temperatures and moderate rainfall (fig. 2). Less intense major flushing periods (between 7.6 ± 0.9 and 61.4 ± 5.7 flushes m\(^{-2}\)) occurred in late spring, mid and late summer, apparently in response to rainfall.

Natural enemies

Spiders and arboreal ants of the subfamily Pseudomyrmecinae were the most frequently encountered...
natural enemies in the stem tap samples ($n = 1312$ and 832, respectively) whereas lacewings and lady beetles were seen only sporadically ($n = 88$ and 79, respectively). Spider abundance varied throughout the year ($F_{7,34.4} = 31.06, P < 0.0001$) and was greatest during autumn and least in spring 2011 (fig. 3a). Pseudomyrmecines also varied significantly over seasons ($F_{1,10.9} = 21.48, P < 0.0007$) and were most abundant in summer (fig. 3b). Numbers of lacewings also varied seasonally ($F_{7,32.4} = 5.22, P = 0.0005$) and were most abundant in winter and least in spring (fig. 3c). Ladybeetle numbers showed no consistent seasonal patterns (fig. 3d) nor significant seasonal variation (Kruskal–Wallis, $H_7 = 10.58, P = 0.1577$ or $H_7 = 5.2375, P = 0.6310$ for unsprayed or sprayed plots, respectively).

Insecticide sprays significantly reduced the cumulative number of spiders observed over all seasons ($F_{1,10.9} = 21.48, P < 0.0007$) and every season except spring 2011 and summer 2012 (fig. 3a). However, the interaction of treatment and season was significant ($F_{7,34.4} = 8.9, P < 0.0001$) indicating seasonal patterns depended on the effect of treatment and vice versa. In general, there was less variation in spider density within treated compared to untreated plots. For instance, the greatest spider incidence in untreated plot occurred during autumn 2011, whereas spider densities were not significantly different in treated plots from summer through autumn 2011 and spring through autumn 2012.

Similar patterns were seen with ant abundance: significant depression of populations in treated plots over all seasons ($F_{1,10.9} = 21.48, P < 0.0007$) and every season except spring 2011 and winter 2012 (fig. 3b). Again, the interaction of treatment and season was significant ($F_{7,35} = 6.72, P < 0.0001$) indicating different seasonal effects depending on the ACP management treatment and vice versa. Plots that did not receive insecticides showed the highest arboreal ant densities in summer and the lowest densities in winter, whereas ‘calendar’ plots did not show much seasonal variation.

No treatment effects ($F_{1,7.64} = 0.31, P = 0.5951$) or treatment interactions with season ($F_{7,32.4} = 0.89, P < 0.5228$) were seen with lacewing abundance (fig. 3c). However, ladybeetle abundance was greater in ‘no insecticide’ plots than in ‘calendar’ plots in autumn 2011, winter 2011, spring 2012, summer 2012 and winter 2012 (fig. 3d).

Fig. 3. Comparison between the two Asian citrus psyllid (ACP) management strategies tested, insecticide ‘calendar’ applications and ‘no insecticides’, in cumulative number per meteorological season of different natural enemies groups (mean±SE) collected by stem tap sampling 20 trees per plot in plots where the exclusion experiments were performed: (a) Araneae, (b) Pseudomyrmecinae, (c) Neuroptera and (d) Coccinellidae. Black columns with the same upper case letter above represent predator abundance in treated plots that are not significantly different. White columns with the same lower case letter above represent predation rates in untreated plots that are not significantly different (LSMEANS $P < 0.05$ for all except Kruskal–Wallis test for Coccinellidae). Asterisks within a white column indicate differences between treatments during the respective season (two stars, $P < 0.05$, one star, $P < 0.10$).
Exclusion experiments

Estimated predation of ACP varied significantly with season ($F_{2,41.3} = 3.98$, $P = 0.0001$). In general, higher predation rates were found in spring and summer compared to late winter within the first flushing period of the season (fig. 4).

Corrected mortality presumed to predation of ACP colonies across all eight repetitions of the experiment ranged between 0 and 87.95 ± 6.28% for the ‘calendar’ treatment and 14.50 ± 9.85 and 91.29 ± 4.42 in untreated blocks, and was significantly higher for the ‘no insecticide’ treatment ($F_{1,4.85} = 22.89$, $P = 0.0053$). The interaction between spray treatment and season was significant ($F_{2,41.3} = 2.76$, $P = 0.0189$) indicating differences in insecticide use depended on seasonal pattern and vice versa. Nevertheless, no significant treatment effects on predation rates were seen until the second year, in February 2012 ($t_{22} = −2.31, P = 0.0306$) and May 2012 ($t_{25} = −3.83, P = 0.0008$) and again in February 2013 ($t_{19.1} = −1.86, P = 0.0779$). Predation rates in the calendar treatment were either very low on these dates or zero on the last date; i.e., no difference between open and closed cages ($F_{1,16} = 2.71, P = 0.1191$).

ACP growth rates

The negative exponential distribution effectively modeled the relationship in ‘no insecticide’ plots between predation rates on the one hand ($r$) and the ACP population growth rate ($\lambda$) obtained from monitoring ACP adult populations before and after major flush cycles ($F_{2,5} = 84.94, P = 0.0001; r^2 = 0.97$) (fig. 5). The model predicted ACP population growth rates of $\lambda = 3.98$ in insecticide treated plots compared to $\lambda = 0.56$ in no insecticide plots based on estimated ACP predation rates ($r$) during the major flushing periods in the first year (table 2). Predicted values for $\lambda$ the second year were much higher, 14.6 and 4.15 for insecticide treated and not treated, respectively.

Discussion

Arboreal spiders were the most frequently encountered predators using stem tap sampling. Spiders are recognized as a key predacious arthropod group in agriculture (Sunderland, 1999). However, their generalist predatory behavior, low numerical response to specific prey and typically long life cycle have generally led to an underestimation of the services they provide in agroecosystems (Riechert & Lockley, 1984; Sunderland, 1999; Symondson et al., 2002). Ecotoxicology studies on this group are scarce, comprising only 3% of the total according to a recent review (Pekar, 2012). We found significant reduction of spiders and an advance of their seasonal activity peaks in response to monthly insecticide treatments (fig. 3a). Effects of insecticides on spiders can depend on active ingredient, targeted guild or species (Mansour et al., 1980; Mansour & Nentwig, 1988; Pekar, 1999; Amalin et al., 2000; Pekar, 2002). Pyrethroids, organophosphates and carbamates can be highly toxic to spiders both in laboratory and field studies (Mansour et al., 1980; Amalin et al., 2000; Fountain et al., 2007). These modes of action were used every fall and winter plus 4 other times in 2011 (table 1) which could explain why spider populations were at their lowest ebb in treated plots compared to untreated plots during the winters of 2011 and 2012. Treatment effects on spiders in winter might also explain reduced predation rates the following spring. In contrast, spirotetramat, spinetoram and diflubenzuron used later in the year are considered non-toxic to spiders, and abamectine spirotetramat, spinetoram and diflubenzuron used later in the year are considered non-toxic to spiders, and abamectine or foliar imidacloprid are reported as only moderately toxic (Hassan et al., 1994; Amalin et al., 2000; Bajwa & Aliniazee, 2001). The use of more selective active ingredients in 2012 could explain the apparent recovery of spider populations in sprayed plots during summer 2012, although species composition could still have remained altered by pesticides (Pekar, 2012).

Arboreal ants of the genus Pseudomyrmex were the second predatory group in abundance. Behavior of these ants is recognized to be almost exclusively predatory (Larsen & Philpott, 2010). In citrus, they are considered potential predators of citrus leafminer (Xiao et al., 2007). However, the importance of arboreal ants as biological control agents has...
been lately ignored despite the early example of the weaver ant *Oecophylla smaragdina* used for biological control on citrus in China (Chen, 1962). To our knowledge, the role of *Pseudomyrmex* spp. as predators of ACP has not been assessed although we have observed workers carrying off ACP nymphs, as did Michaud (2004). In our study, insecticide sprays had a strong negative effect in population of these ants almost from the onset of treatments. There are no published reports of pesticide side effects on this group, but most of the studies in southwest Florida with no survivorship observed in unsprayed areas. However, it is also possible that intensive area-wide spraying for psyllids has resulted in significant depression of lady beetle populations. A reduction of the lady beetle metapopulations would delay re-colonization processes even in unsprayed areas.

We observed differences in survivorship from spring to fall of 50–90% between caged and uncaged cohorts of *D. citri* that were presumably due to predation. These results are in concert with previous studies that documented the importance of predation on nymphs as a key source of mortality inflicted on *D. citri* populations (Michaud, 2001, 2004; Qureshi & Stansly, 2009). The estimated reduction in net ACP reproductive rate (*R₀*) ranged from 3- to 178-fold over 16 cohort exclusion studies in southwest Florida with no survivorship observed in the 17th (June) experiment (Qureshi & Stansly, 2009). Unfortunately, this mortality was insufficient to stop the rapid spread of HLB in the region, necessitating insecticidal control (Qureshi & Stansly, 2008, 2010). Nevertheless, it is important to assess the impact of different insecticidal programs on this natural enemy component as a step toward integration of biological and chemical control.

The calendar insecticide program we used for ACP management made minimal use of broad-spectrum insecticides in an effort to conserve natural enemies. Still, it caused significant reductions of predation rates to ACP cohorts during February both years and in May 2012. Super-abundance of young flush in late winter provided opportunity for rapid growth of the ACP population (high λ) where biological control was negatively impacted by insecticides, an effect which carried into late spring in 2012 (table 2). In contrast, no significant treatment differences in presumed predation rates were observed during summer or autumn in spite of significant reduction of key natural enemy population by insecticides during that time (fig. 3a–d), possibly due to generally low finite rates of increase (λ) of ACP due to lack of flush during those periods (fig. 1, table 2). While the system has proven itself resilient to insecticidal insult over the short term, continuous use is likely to take its toll over medium and long-term.
longer terms as indicated by generally lower predation rates observed in this compared to a previous study (Qureshi & Stansly, 2009). The cumulative effect of spraying could also be seen in the latter study, where no effects were seen on ACP predation until spring 2012 following a year of monthly sprays.

In conclusion, 2 years of an intensive insecticide program for ACP management resulted in a reduction in abundance and in some cases a modification of seasonal patterns within some key predatory groups of the citrus agroecosystem. Deleterious effects of insecticides on citrus predatory assemblages had direct consequences on the natural occurring biological control of ACP early in the growing season when spring flush provided the greatest opportunity for population growth. At that time, even small differences in biological control could translate into rapid ACP population increase. Dormant season sprays have been shown to suppress overwintering populations of ACP with minimal impact on key natural enemies (Qureshi & Stansly, 2010). While this allows predation to help control ACP later in the year, direct or sublethal effects of continuing sprays may reduce functionality of natural enemy populations in the medium- or long-term (Desneux et al., 2007; Biondi et al., 2012). The results obtained in this study may therefore underestimate the negative consequences of insecticides on beneficial arthropod populations over a long-term scenario, and highlight how fewer and more selective insecticide applications during critical periods could be favorable for overall ACP management.

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